

International Journal of Agricultural Extension and Rural Development Vol. 8 (8), pp. 001-005, August, 2020. Available online at www.internationalscholarsjournals.org © International Scholars Journals

Author(s) retain the copyright of this article.

Full Length Research Paper

Brassica oleracea genotypes displaying interesting fatty acid profiles for Brassica napus breeding

F. Seyis^{1,2}* and W. Friedt²

¹Department of Field Crops, Faculty of Agriculture, Bozok University, Yozgat, Turkey. ²Institute of Plant Breeding, Justus-Liebig-University, Giessen, Germany.

Accepted 22 June, 2019

Brassica oleracea (CC) normally displays an erucic acid (22:1) content ranging from 28 to 63%. In the course of studies dealing with the inheritance of erucic acid content in the seed oil of Brassica species individual plants belonging to three accessions of B. oleracea conv. capitata - Kashirka 202, Ladozhskaya DS 8395 and Eisenkopf - were identified displaying low, intermediary and high erucic acid content. The fatty acid profiles of these cabage genotypes and their potential use in Brassica napus breeding is discussed.

Key words: Brassica oleracea, cabbage, low-erucic acid mutant, fatty acid composition, breeding.

INTRODUCTION

Rapeseed and mustards derived from several locally distributed members of the genus Brassica have become one of the worldwide most important sources of vegetable oil due to substantial progress in breeding and cultivation practice. Especially Brassica napus displaying 'double-low' seed quality (low erucic acid, low glycosino-late content) dominates field crop production in several European countries with cool-temperate climates. Efforts to develop low erucic character in several Brassica species led to the discovery of low-erucic acid mutants in the species Brassica rapa (AA) (Downey, 1964), B. napus (AACC) (Stefansson et al., 1961; Stefansson and Hougen, 1994) and Brassica juncea (AABB) (Kirk and Oram, 1978). In amphidiploid B. carinata (BBCC) low-erucic acid mutants were not known up to the 90's. Different strategies, such as the selection of trans-gressive segregants in the crossing progeny of diverse B. carinata accessions (Alonso et al., 1991), induced mutations and interspecific crosses with low-erucic B. napus and B. juncea (Getinet, 1987; Fernandez-Escobar et al., 1988) were used to reduce the erucic acid content. With the first and last mentioned method low erucic acid forms were selected successfully (Alonso et al., 1991; Getinet et al., 1994). For the other monogenomic species

Brassica oleracea (CC) and B. nigra (BB) low-erucic acid forms were not cited in the literature up to now (Downey, 1964; Stefansson et al., 1961; Stefansson and Hougen, 1994).

In the course of studies dealing with the inheritance of erucic acid content in the seed oil of Brassica species, cabbage genotypes displaying low erucic character were identified. Individual plants belonging to the three B. oleracea accessions, namely Kashirka 202 Ladozhskaya and Eisenkopf, were identified being very low in erucic acid content. The fact that all these genotypes display low erucic acid content and that a monogemic inheritance was detected in Kashirka were published before (Lühs et al., 2000; Seyis et al., 2004). During the mentioned studies dealing with the genetics of erucic acid content individual half seed plants with intermediary and high erucic acid were also determined besides these half seed plants displaying low erucic acid content. Between the well known six Brassica species, oilseed rape (Brassica napus L.) is the most important agricultural crop plant. Rapeseed is an interspecific amphidiploid hybrid between two unknown parents from the diploid species B. rapa and B. oleracea. But the genetic variation in the present gene pool of B. napus is rather limited. The reason for this limited trait variation found within the B. napus genetic resources available for breeding is the result of oilseed rape's relatively recent origin from only two parents and from the intensively quality breeding of this plant in the last decades.

^{*}Corresponding author. E-mail: fatih.seyis@bozok.edu.tr. Tel: +90-354-2123577. Fax: +90-354-2122789.

Interspecific hybridization has great potential for the improvement of Brassica crops (Inomata, 1997) as it creates genetic variation (Prakash, 1973; Choudhary, 1997) and is a valuable tool for transferring traits from one species to another (Prakash and Chopra, 1988; Raney et al., 1995a, b). Therefore, one way of increasing trait variation is the development of so called resynthesised rapeseed forms using the diploid progenitors.

The aim of modern plant breeders are to develop higher yielding, nutritious and environmentally friendly varieties that improve our quality of life without harnessing additional natural habitats to agricultural production (Zamir, 2001). Without a broad base of heterogeneous plant material, it is impossible for plant breeders to produce cultivars that meet the changing needs regarding adaptation to growing conditions, resistance to biotic and abiotic stresses, higher product yield or specific quality requirements (Friedt et al., 2007). Therefore, the most efficient way to improve the performance of crop varieties further is to have access to a large diverse pool of genetic diversity. Especially, interspecific hybridizations are a useful tool for the broadening of genepools. Considering, that B. napus is an amphidiploid plant and that the genetic diversity in B. oleracea is very rich, the use of resynthesised rapeseed forms will broaden the present narrow genetic basis in the B. napus gene pool.

Especially the development and selection of resynthesised rapeseed genotypes displaying low erucic acid character is important, because the low erucic character present in the gene pool of *B. napus* has an intraspecific origin. Canadian plant breeders were identified plants with low eicosenoic acid and erucic acid content (Downey and Harvey, 1963; Stefansson et al., 1961) in spring rapeseed cultivars. The potential use of *B. oleracea* genotypes, described regarding their fatty acid composition, in *B. napus* breeding is discussed.

MATERIALS AND METHODS

Plant matarial

Seed material was taken from the USDA-ARS Plant Genetic Resources Unit, Cornell University, Geneva, New York, USA.

Methods

The preparation of half seeds of the genotypes Kashirka and Ladozhskaya and Eisenkopf was done according to the method described by Lühs (1996) and Thies (1971).

RESULTS

During these experiment half seed plants displaying different erucic acid contents would be determined and these half seed plants were divided regarding their fatty acid acid composition in three groups (Table 1). These

groups are respectively the low erucic acid group (0 - 3% erucic acid), the intermediary group (13 - 39%) and the high erucic acid group (>40% erucic acid). The numbers of investigated half-seed plants for every genotype in group are given in brackets (Table 1). In the zero erucic acid groups, the amount for every fatty acid seem to be very similar for all genotypes regarding mean values (Table 1).

The highest oleic acid content was determined in Kashirka (57,46%), followed by Eisenkopf (56,51%) and Ladozshkaya (55,35%). Correspondingly, the erucic acid content of all genetypes was low. In the intermediary group Kashirka and Eisenkopf showed higher amounts of erucic acid (C22:1) compared with Ladozhskaya, respectively 28,74% and 30,43%. Ladozhskaya display higher amounts of oleic acid (29,26%), but lower amounts of erucic acid (18,8%). In the high erucic acid group, Ladozhskaya and Eisenkopf showed higher erucic acid contents (both 45,1%), followed by Kashirka (42,7%).

DISCUSSION

Contrary to oilseed rape and turnip rape (Lühs et al., 2000; Jönsson, 1977; Anand and Downey, 1981; Chen and Heneen, 1989; Lühs and Friedt, 1995a) studies regarding fatty acid composition of *B. oleracea* (CC) seed oil are lacking (Lühs et al., 2000). This rather neglected *Brassica* species displays normally a high erucic acid (C22:1) content ranging between 28 - 63% (Lühs and Friedt, 1995b). Previous studies indicated an additive mode of inheritance in some *Brassicaceae* species including *B. rapa* (Dorrell and Downey, 1964), *B. napus* (Harvey and Downey, 1964; Chen and Heneen, 1989; Lühs and Friedt, 1995a), *B. juncea* (Kirk and Hurlstone, 1983), *B. carinata* (Getinet et al., 1994) and *Sinapis alba* (Raney et al., 1995).

In a mentioned previous study after applying half-seed selection, the subsequent analysis of the progeny might confirm whether erucic acid content is determined by one gene locus as one could expect (Lühs et al., 2000). So, it could be detected that the genotype Kashirka displays a monogenic inheritance regarding erucic acid content. Due to the different erucic acid distribution seen in Ladozhskaya in the intermediary group it cannot be said that there is a monogenic inheritance, but this needs further detailed investigation.

The discovery, that *B. napus* is an interspecific hybrid (resynthesised rapeseed = RS rapeseed) of the cross between the diploid *Brassica* species *B. oleracea* and *B. rapa* (Morinaga, 1934; U, 1935) and the determination that resynthesised rapeseed forms are representing genetically a new genepool compared with present breeding material (Song et al., 1993; Seyis et al., 2003a; Voss et al., 1998; Becker et al., 1995), give reason for the idea to use resynthesised rapeseed forms in producing either semisynthetic hybrids through crossing with

Table 1. Fatty acid composition in mean values of investigated individual *Brassica oleracea* plants (number of analyzed plants are given in brackets).

Genotype		C16:0	C18:0	C18:1	C18:2	C18:3	C20:1	C22:1	C24:1
Zero erucic	Kashirka (n = 24)	3.58	0.94	57.46	22.07	12.93	1.90	0.21	0.28
acid type	Min-MaxValues	3-1-4.4	0.2 - 1.3	53.2 - 63.4	17.7 - 26.5	11.2 - 15.6	1.5 - 2.5	0-0.6	0.1- 0.6
	Standart Deviation	0.37	0.22	2.85	2.42	1.3	0.25	0.19	0.13
	Ladozshkaya (n = 25)	3.92	1.09	55.35	21.24	14.56	2.81	0.62	0.32
	Min-MaxValues	3.1 - 4.8	0.7-2.1	38.9 - 65.5	16.0 - 24.7	11.3 - 23.0	0-8.5	0-2.4	0-0.9
	Standart Deviation	0.5	0.3	6.8	4.2	2.9	2.7	0.7	0.2
	Eisenkopf (n = 20)	5.84	1.09	56.51	23.63	11.48	1.76	0.36	0.30
	Min-MaxValues	4.4 -7.5	0.7-1.4	46.5 - 65.5	15.0 - 32.1	8.1 - 14.4	0.4 - 5.0	0-1.2	0.1- 0.7
	Standart Deviation	8.0	0.2	5.9	5.9	1.5	1.0	0.4	0.2
Intermediary	Kashirka (n = 21)	2.97	0.88	24.77	13.96	11.23	15.78	28.74	0.68
type	Min-MaxValues	2.6 - 3.7	0.6-1.2	21.5 - 28.9	10.5 - 17.2	9.5-14.4	12.9 - 21.0	20.8 - 34.8	0.4 - 1.4
	Standart Deviation	0.3	0.1	2.2	1.7	1.2	2.1	3.9	0.3
	Ladozshkaya (n = 33)	3.40	0.89	29.26	15.78	11.76	12.11	24.56	1.24
	Min-MaxValues	2.6 - 4.4	0.6 - 1.4	18.5 - 39.3	9.0-21.9	8.2-17.3	8.6 - 16.4	13.3 - 38.3	0.1 - 2.3
	Standart Deviation	0.5	0.3	1.7	2.7	2.1	2.5	9.5	0.6
	Eisenkopf (n = 33)	4.48	0.96	23.38	15.41	9.64	12.95	30.43	1.41
	Min-MaxValues	2.5 - 6.9	0.6 - 1.6	14.4 - 30.8	10.3-22.2	0.00-17.0	10.3 - 16.4	23.4-39.6	0.4 - 2.7
	Standart Deviation	1.0	0.2	4.0	3.2	2.8	1.6	4.5	0.5
High erucic acid type	Kashirka (n = 3)	2.90	0.67	19.23	10.43	9.53	12.03	42.70	1.05
	Min-MaxValues	2.4 - 3.2	0.6 - 0.8	18.3 - 20.7	9.50-12.2	8.5 - 10.8	10.1-14.8	40.4 - 47.2	0.5 -1.2
	Standart Deviation	0.44	0.2	1.29	1.53	1.17	2.46	3.9	0.35
	Ladozshkaya (n = 12)	2.77	0.67	15.93	12.51	10.58	9.90	45.1	1.31
	Min-MaxValues	2.0 - 3.6	0.6 - 0.8	12.1-19.4	9.0 - 14.2	7.6 - 13.5	7.9 - 11.6	36.3 - 52.5	0.6 - 1.9
	Standart Deviation	0.49	0.1	2.94	1.55	1.82	1.44	4.16	0.41
	Eisenkopf (n = 22)	3.41	0.90	15.2	12.4	9.5	10.10	45.1	1.5
	Min-MaxValues	2.0 - 6.5	0.62 - 1.2	12.4 - 19.1	8.9 - 21.7	6.6 - 13.3	7.7 - 12.5	40.3 - 52.2	1.0 - 2.6
	Standart Deviation	1.12	0.19	1.9	3.4	1.86	1.38	2.82	0.5

conventional material or in developing true hybrids through crossing with msl-lines (msl = Männliche Sterilität Lembke) (Seyis et al., 2001; Seyis et al., 2003b; Lühs et al., 2003). For this purpose low erucic acid mutants of *B. oleracea* were crossed

with double low *B. rapa* forms to develop resynthetic rapeseed with low erucic acid content (Lühs et al., 2003; Seyis et al., 2005). Success in removal of erucic acid led many plant breeders to look at other possible modifications in rapeseed.

One of the first objectives to increase the level of erucic acid in order to increase the value of oil used for the industrial production of chemicals (Latta, 1990; Sonntag, 1991; Lühs, 1996). Several strategies have been developed (Taylor et al.,

1992) but there has been no reports of cultivars with erucic acid levels (HEAR) greater than 55% to date. Beside HEAR and LEAR (Low erucic acid rapeseed) several other fatty acid modification types have been reported in the literature, but do not seem to be under commercial production – these include types with high or low levels of saturated fatty acids (Persson, 1985; Friedt and Lühs, 1998; Thelen and Ohlrogge, 2002) as well as alcolohols and waxes (Anonymous., 2003). Some of the delay in producing commercialized varieties of these types has been the general resistance towards the acceptance of genetifically modified oils (Ratnayake and Daun, 2002).

The three investigated B. oleracea accessions display interesting fatty acid profiles. With the use of these genotypes in interspecific crosses with different quality type B. rapa forms it will be possible to create intersprecific rapeseed forms with different fatty acid compositions, which are different from present rapeseed genetic material, without using genetic transformation. Further, the mentioned comparatively narrow genetic basis in rapeseed could be increased with the use of the both highly polymorphic diploid parents, what offer a much broader genetic variability that then can be exploited. In general, the low yield performance and conventional quality (high erucic acid-high glucosinolates rapeseed, HEAR) of resynthesised B. napus is a handicap for the broad use of this novel genepool in modern rapeseed breeding programmes. However, with the use of low erucic acid mutants among B. oleracea accessions and the development of synthetic rapeseed through wide crosses with respective 0- or 00-quality B. rapa genotypes will offer the possibility to use this basic material as a genetic resource for quality and yield improvement in oilseed rape breeding.

REFERENCES

- Anonymous (2003). Improved canola holds the key to the future. Canola Digest p. 22.
- Alonso LC, Fernadez-Serrano O, Fernadez-Escobar J (1991). The outset of a new oilseed crop: *Brassica carinata* with low erucic acid content. Proceedings 8th Int. Rapeseed Congress (GCIRC), Saskatoon, Sask. Canada: pp. (1), 170-176.
- Anand IJ, Downey RK (1981). A study of erucic acid alleles in digenomic rapeseed (*Brassica napus* L.). Can. J. Plant Sci., 61: 199-203.
- Becker HC, Engqvist GM, Karlsson B (1995). Comparison of rapeseed cultivars and resynthesized lines based on allozyme and RFLP markers. Theor. Appl. Genet., 91: 62-67.
- Choudhary BR (1997). Interspecific hybridization in genus Brassica. PhD Thesis, Rajasthan Agricultural University, Bikaner, India.
- Chen BY, Heneen WK (1989). Fatty acid composition of resynthesised *Brassica napus* L., *B. campestris* L. and *B. alboglobra* Bailey with special reference to the inheritance of erucic acid content. Heredity, 63: 309-314.
- Dorrell DG, Downey RK (1964). The inheritance of erucic acid content in rapeseed (*Brassica campestris*). Can. J. Plant Sci., 44: 499-504.
- Downey RK (1964). A selection of *Brassica campestris* L. containing no erucic acid in its seed oil. Can. J. Plant Sci., 44: 499-504.
- Downey RK, Harvey BL (1963). Methods for breeding for oilseed quality in rape. Canadian J Plant Sci., 43: 271–275.

- Fernandez-Escobar J, Dominguez J, Martin A, Fernadez-Martinez JM (1988). Genetics of the erucic acid content in interspecific hybids of Ethiopian Mustard (*Brassica carinata* Braun) and rapeseed (*B. napus* L.). Plant Breeding, 100: 310-315.
- Friedt W, Lühs W (1998). Recent developments and perspectives of industrial rapeseed breeding. Fett/Lipid, 100: 219-226.
- Friedt W, Snowdon RJ, Ordon F, Ahlemeyer J (2007). Plant Breeding: Assessment of Genetic Diversity in Crop Plants and its Exploitation in Breeding. Progress in Botany, 648: 151-178.
- Getinet A (1987). Review on breeding of Ethiopian mustard (*Brassica carinata* A. Braun). Proc 7th Inter. Rapeseed Congress (GCIRC), Poznan, Poland, 2: 593-597.
- Getinet A, Rakow G, Raney JP, Downey RK (1994). Development of zero erucic acid Ethiopian mustard through an interspecific cross with zero erucic acid oriental mustard. Can. J. Plant Sci., 74: 793-795.
- Harvey BL, Downey RK (1964). The inheritance of erucic acid in rapeseed (*Brassica napus*). Can. J. Plant Sci., 44: 104-111.
- Inomata N (1997). Wide hybridization and meiotic pairing. In: Kalia HR, Gupta SK (ed.) Recent advances in oilseed Brassicas. Kalyani Publ, Ludhiana, pp. 53–57.
- Jönsson R (1977). Erucic acid heredity in rapeseed (*Brassica napus* L. and *Brassica campestris* L.). Hereditas, 86: 159-170.
- Kirk JTO, Hurlstone CJ (1983). Variation and inheritance of erucic acid content in *Brassica juncea*. Z. Pflanzenzüchtg, 90: 331-338.
- Kirk JTO, Oram RN (1978). Mustards as possible oil and protein crops for Australia. J. Austr. Inst. Agric. Sci., 44: 143-156.
- Latta S (1990). New industrial uses of vegetable oils. Informatics, 1: 434–443.
- Lühs W, Friedt W (1995a). Breeding high-erucic acid rapeseed by means of *Brassica napus* resynthesis. In: Proc. 9th Int. Rapeseed Congr., Cambridge, UK, 2: 449-451.
- Lühs W, Friedt W (1995b). Natural fatty acid variation in the genus *Brassica* and its exploitation through resynthesis. Eucarpia Cruciferae Newslett. 17: 14-15.
- Lühs W (1996). Genetic-analytical investigations for breeding rapeseed (*Brassica napus* L.) with maximal erucic acid content in ist seed oil as industrial raw material. University of Giessen. Niedekleen/Hesse, Wissenschaftl. Fachverlag Dr Fleck, 164 pp.
- Lühs W, Seyis F, Voss A, Friedt W (2000). Genetics of erucic acid content in *Brassica oleracea* seed oil. Czech. J. Genet. Plant Breed. 36: 116-120.
- Lühs W, Seyis F, Baetzel R, Friedt W (2003): Genetic diversification of Brassica napus seed quality by wide hybridisation. 11th Intern. Rapeseed Congr., 6-10 July 2003, Copenhagen, Denmark, 2: 375-377.
- Morinaga T (1934). Interspecific hybridization in *Brassica*. VI. The cytology of F1 hybrids of *B. juncea* and *B. nigra*. Cytologia (Tokyo), 6: 62-67
- Prakash S (1973). Artificial synthesis of Brassica juncea Cross. Genetica, 44:2 49–263.
- Prakash S, Chopra VL (1988). Synthesis of alloplasmic Brassica campestris as a new source of cytoplasmic male sterility. Plant Breeding, 101: 253-255.
- Persson C (1985). High palmitic acid content in summer turnip rape (*Brassica campestris* var. Annua L.). Cruciferae Newslett., 10: 137.
- Raney P, Rakow G, Olson T (1995). Development of low erucic, low glucosinolate *Sinapis alba*. Proc. 9th Int. Rapeseed Congr., Cambridge, UK. 2: 416-418.
- Raney P, Rakow G, Olson T (1995a). Modification of Brassica seed oil fatty composition utilizing interspecific crossing. In: Proc 9th Int Rapeseed Congr, vol 2, Cambridge, UK, pp. 410–412.
- Raney P, Rakow G, Olson T (1995b). Development of zero erucic, low linolenic Brassica juncea utilizing interspecific crossing. In: Proc 9th Int Rapeseed Congr, vol 2, Cambridge, UK, pp. 413–415.
- Raney P, Rakow G, Olson T (1995c). Development of low erucic, low glucosinolate Sinapis alba. Proc. 9th Int. Rapeseed Congr., Cambridge, UK. 2: 416-418.
- Ratnayake WMN, Daun JK (2002). Chemical composition of canola and rapeseed oils. In: Rapeseed and Canola Oil. Ed. F. D. Gunstone, Blackwell Publishing Limited, Oxford (UK) 2004, pp. 37–73.
- Song K, Tang K, Osborn TC (1993). Development of synthetic Brassica amphidiploids by reciprocal hybridization and comparison to natural

- amphidiploids. Theor. Appl. Genet., 86: 811-821.
- Sonntag NOV (1991). Erucic, behenic: feedstocks of the 21st century. INFORM, 2: 449–463.
- Seyis F, Friedt W, Lühs W (2001). Resynthese-Raps (*Brassica napus* L.) als genetische Ressource für die Qualitäts- und Ertragszüchtung.
 In: K. Hammer und T. Gladis (Hrsg.), Nutzung genetischer Ressourcen ökologischer Wert der Biodiversität. Schriften zu Genetischen Ressourcen, 16: 91-112.
- Seyis F, Snowdon RJ, Lühs W, Friedt W (2003a). Molecular characterization of novel resynthesized rapeseed (*Brassica napus*) lines and analysis of their genetic diversity in comparison with spring rapeseed cultivars. Plant Breeding, 122: 473-478.
- Seyis F, Friedt W, Lühs W (2003b). Resynthesised *Brassica napus* as genetic resource in rapeseed improvement for quality and agronomic performance. In: H. Knüpffer and J. Ochsmann (eds.), Rudolf Mansfeld and Plant Genetic Resources. Proceedings of a symposium dedicated to the 100th birthday of Rudolf Mansfeld, Gatersleben, Germany, 8-9 October 2001. Schriften zu Genetischen Ressourcen, 19: 336-340. ZADI/IBV, Bonn.
- Seyis F, Friedt W, Voss A, Lühs W (2004). İdentification of İndividual *Brassica oleracea* plants with low erucic acid content. Asian J. Plant Sci., 3(5): 593-596.
- Seyis F, Friedt W, Lühs W (2005). Development of Resynthesized Rapeseed (*Brassica napus* L.) Forms with Low Erucic Acid Content Through *in ovulum* Culture. Asian J. Plant Sci., 4(1): 6-10.

- Stefansson BR, Hougen FW, Downey RK (1961). Note on the isolation of rape plants with seed oil free from erucic acid. Can. J. Plant Sci., 41: 218-219.
- Stefansson BR, Hougen FW (1964). Selection of rape plants (*Brassica napus*) with seed oil practically free from erucic acid. Can. J. Plant Sci., 44: 359-364.
- Taylor DC, Barton DL, Rioux KP, MacKenzie SL, Reed DW, Underhill EW, Pomeroy MK, Weber N (1992). Biosynthesis of acyl lipids containing very-long chain fatty acids in microspore-derived embryos of *Brassica napus* L. cv. Reston. Plant. Physiol., 99: 1609-1618.
- Thelen JJ, Ohlrogge JB (2002). Metabolic Engineering of Fatty Acid Biosynthesis in Plants. Metabolic Eng., 4 (1): 122-21.
- Thies W (1971). Rapid and easy analysis of fatty acid composition in individual rapeseed cotyledons. Z. Pflanzenzüchtg. 65: 181-202.
- U N (1935). Genome analysis in *Brassica* with special reference to the experimental formation of *B. napus* and peculiar mode of fertilization. Jap. J. Bot., 7: 389-452.
- Voss A, Friedt W, Marjanovic-Jeromela A, Lühs W (1998). Molecular genotyping of rapeseed including resynthesized *Brassica napus* lines. Cruciferae Newslett., 20: 27-28.
- Zamir D (2001). Improving plant breeding with exotic genetic libraries. Nat. Rev. Genet., 2: 983-989.