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Full Length Research Paper

Physicochemical Characterization of Nitisol in Southwestern Ethiopia and Its Fertilizer Recommendation Using NuMaSS

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Nutrient management decision support system (NuMaSS) is a tool that diagnoses soil constraints and selects the appropriate management practices, based on agronomic, economic and environmental criteria, for location-specific conditions. However, its validation in different soil types and agroecology should be tested. Thus, the present study was conducted to assess the fertility status of Nitisol and suggests the application of NuMaSS as a diagnostic tool to recommend supplementary N and P fertilizers for maize across different agroecology zones. Accordingly, fifty soil samples were collected from five locations and analyzed for the major soil physical and chemical properties. The soil pH-H₂O was below the critical level for all soil samples. Nitrogen, Cu and Zn were deficit in all soil samples. Deficiency of available phosphorous was also observed in 88% of the soil samples. High probabilities of nitrogen and phosphorous deficiency were also observed in the study areas using NuMaSS. Moreover, 24 Kg/ha P and 73 Kg/ha N for Dedo; 24Kg/ha P and 82 Kg/ha N for Agaro; 20 Kg/ha P and 100 Kg/ha N for Serbo; 27 Kg/ha P and 94 Kg/ha N for Dimtu; and 4.9Kg/ha P and 97 Kg/ha N for Asendabo were recommended for maize using NuMaSS.

Keywords: NuMaSS, Nitisol, Soil fertility, Critical level, Deficiency

INTRODUCTION

Declining of soil fertility is a fundamental impediment to agricultural growth and a major reason for slow growth in food production in Sub-Saharan Africa (SSA) (Sánchez et.,al 1995; Quinones et al., 1998; Shapiro and Sanders, 1998). Moreover, low soil fertility is recognized as an important constraint to increased food production and farm incomes in many parts of sub-Saharan Africa (Shepherd and Soule, 1998). In

Ethiopia, soil degradation and nutrient depletion have gradually increased and have become serious threats to agricultural productivity (Hurni, 1988; Campbell et al., 1996; Bojo and Cassels, 1995; Fassil and Charles, 2009). Nutrient balance calculations for countries by Stoorvogel and Smaling (1990) also showed that Ethiopia was among the countries with the highest rates of net nutrient losses. The annual nutrient deficit

is estimated at -41 kg N, -6 kg P and -26 kg K ha . Thus, there is an urgent need to improve soil fertility and nutrient management in the country.

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Table 1: Detail description of the study sites

Location	Altitude (m)	Coordinates	Mean	Annual	Mean	Annual
			Rainfall (m	m)	Temperature (⁰ 0	C)
Dimtu	2166	7° 58' 0" N, 37°11' 0" E	1168.00		27.6	
Dedo	2000	7° 31' 0" N, 36°52' 0" E	1500.00		20-25	
Asendabo	1750	7° 53′ 0″ N, 35°25′ 0″ E	1131.08		27.6	
Agaro	1716	7°49'N 36°39'E	1663.00		19.9	
Serbo	1640	7°35' N, 36°46'-37°14'E	1150.00		18-22	

Source: BPEDORS, 2000

Depletion of soil nutrient is a reversible constraint as long as soil test based fertilizer application is in place (Fassil and Charles, 2009). However, assessing soil fertility status is difficult because most soil chemical properties either change very slowly or have large seasonal fluctuations; in both cases, it requires longterm research commitment (Taye and Yifru, 2010). Moreover, the knowledge requirements to properly diagnose and prescribe best management alternatives for location-specific nutrient problems throughout the world exceed the capacity of any human expert. Scarcity and cost of experts; however, can be alleviated by adopting a new system called nutrient management decision support system (NuMaSS). NuMaSS is a tool that diagnoses soil constraints and selects the appropriate management practices, based on agronomic, economic and environmental criteria, for location-specific conditions.

In Jimma zone, Nitosols are the dominant soil types and it is much utilized for crop production. There are about 6827 Km² of Nitisol in the zone covering about 35% of the district's landmass (BPEDORS, 2000). According to Weigel (1986) and Elias (2002), Nitisols are inherently fertile, but large areas in Ethiopia have now been depleted due to continuous cultivation, leaching and erosion. However, the fertility status of Nitisol that dominates the study area is not quantified. usually Furthermore, farmers apply blanket recommendation of fertilizer in the study area irrespective of soil types and agroecology. Therefore the objective of the study is to evaluate the fertility status of Nitisol in Jimma zone and to determine the recommended amount of N and P fertilizer for maize across different locations using NuMaSS.

MATERIALS AND METHODS

Description of the Study Areas

The study was conducted in the Jimma Zone, southwestern Ethiopia, which is located between 7°13′ – 8°56′N latitude and 35°49 -38°38′E longitude with an

estimated area of 19,506.24 km² and the elevation ranges between 1000-3500m. The zone constitute three major climates 78%, 12% &10% belongs to subtropical, temperate and tropical or thermal zones, respectively. The major soil groups of the zone are Nitosol 6827 Km² (35%), Vertisol, 2925.9Km² (15 %) and others 9553 Km² (50%). Almost 49.6% of the zone total area devote to cultivation. The remaining 50.4% of the total area of land were under vegetation cover (22.8% under forest, 18% under woodland, and 9.6% under grassland (BPEDORS, 2000). For the present study, five study areas which have good agricultural potential and dominated by Nitisol were selected from different locations. The detail description of the study areas are presented in Table 1.

Insert table 1 here

Soil Sampling and Analysis

Prior to soil sampling, soil profile studies were carried out in order to characterize and classify the dominant soil types in the study areas using WRB (2006). Accordingly, fifty soil samples were collected randomly from different farmlands at the depth of 25cm. The soil samples were air dried, mixed well, ground, and passed through 2 mm sieve and the laboratory analysis were conducted using the following standard procedures. Soil texture was determined by hydrometer. Soil pH-H2O and pH-KCI were measured by using a pH meter in a 1:2.5 soil: water and soil: KCl respectively. Soil organic carbon was determined by the Walkley-Black oxidation method, total nitrogen by the micro-Kjeldahl digestion, distillation and titration method, and available phosphorus was determined using Bray I extraction method as described by Van Reeuwijk (1992). Total exchangeable bases were determined after leaching the soils with ammonium acetate. Amounts of Ca2+ and Mg²⁺ in the leachate were analyzed by atomic absorption spectrophotometer and K⁺ and Na⁺ were analyzed flame photometrically. Cation exchange capacity was determined at soil pH level of 7 after

Table 2: Mean values of soil separate, pH, EC, exchangeable acidity, OC, TN and available phosphorous across different locations in Jimma zone

Location	Dimtu	Dedo	Asendabo	Agaro	Serbo	Critical	P-value
						level	
Clay (%)	63.33 ^a	41.34 ^b	58.67 ^a	44.67	50.08 ^{ab}	-	*
Silt (%)	23.67 ⁶	34.33 ^{ab}	28.33 ⁰⁰	32.00 ^{ab}	37.92 ^a	-	*
Sand (%)	13.00 ⁰	24.33 ^a	13.00 ⁰	23.33 ^a	12.00 ⁰	-	*
Textural class	Clay	Clay 5.48 ^{au}	Clay	Clay 5.23	Clay		-
pH-H ₂ O	Clay 5.60 ^a	5.48 ^{ab}	5.57 ^a	5.23	Clay 5.35	6.5-8.5	**
pH-KCI	4.39 ^{ab}	4.37 ^{ab}	4.45 ^a	4.28 ⁰	4.04 ^c	-	**
EC (mmhos)	0.050 ^{DC}	0.043 ^c	0.068 ^a	0.057 ⁰	0.046	-	**
Ex.Ac (meq/100g)	b 0.20	0.25 ⁶	b 0.15	0.30 b	0.80 ^a	_	**
OC (%)	2.20	2.71 ^a	1.88 ^c	2.39 ^{ab}	1.50 ^u	0.5-1.0	**
TN (%)	0.18 ^c	0.25 ^a	0.17 ^C	0.22 ^b	0.16 ^c	<1.00	**
C/N	12.10	11.17	10.83	10.96	9.72	10-20	NS
AVP (ppm)	6.63 ^b	6.97 ⁰	8.96 ^a	6.69 ⁰	7.33 ^{ab}	10-15	*

Means in the same row followed by the same letters are not significantly different at 5% level of significance; *significant at P<0.05; **significant at P<0.01; NS = non significance; EC = electrical conductivity; Ex.Ac = Exchangeable acidity; OC = organic carbon; TN = total nitrogen; C/N = carbon to nitrogen ratio; AVP = available phosphorus

displacement by using 1N ammonium acetate method in which it was, thereafter, estimated titrimetrically by distillation of ammonium that was displaced by sodium (Chapman, 1965). Percent base saturation was calculated by dividing the sum of the base forming cations (Ca, Mg, Na and K) by the CEC of the soil and multiplying by 100. Available micronutrients (Fe, Mn, Zn, and Cu) were extracted with DTPA as described by Lindsay and Norvell (Lindsay and Norvell, 1978).

Statistical Analysis

The data were analyzed using descriptive statistics and analysis of variance using SAS 9.2. Duncan's multiple range was also employed to test the significance difference between locations. Correlation analysis was also performed in order to determine the relationship between the selected soil properties. The recommended amounts of nitrogen and phosphorous fertilizers for maize across different locations were determined using Nutrient Management Support System (NuMaSS) version 2.2.

RESULTS AND DISCUSSION

Soil texture

The textural class of Nitisol across all locations was clay; however, there was high variation in soil separates between the sampled soils. The coefficients of variation between the soil samples were 19.70%, 21.06% and 40.49% for clay, silt and sand contents, respectively. Significance difference (P<0.05) in the percentage of soil separates across locations was also

observed (Table 2). All of the soil samples have clay content more than 30%, which is the marginal range of total clay requirement for Nitisol (WRB, 2006). The highest percentages of clay, silt and sand were observed at Dimtu, Sebo and Dedo, respectively while the lowest values were recorded at Dedo, Dimtu and Asendabo, respectively (Table 2).

Soil pH, EC and exchangeable acidity

The statistical analysis showed that soil pH-H₂O was varied significantly (P<0.01) across locations (Table 2). Soil pH-H₂O ranged from 5.23 at Agaro to 5.60 at Dimtu. All of the soil samples had pH-H₂O less the critical level (6.5-8.5) given by Landon (1991). From the sampled soils, 16% rated as strongly acidic while the rest rated as moderately acidic. According to Landon (1991) and Tisdale et al. (1993), most nutrients for field crops are available at pH value of above 5.5; however, 67% of the sampled soils have pH less than 5.5. The lowest values of soil pH at the study sites could be because of loss of base forming cations down the soil profiles even beyond sampling depth through leaching and drain to streams in runoff generated from accelerated erosion. This enhances the activity of Al³⁺ and H in the soil solution, which reduces soil pH and thereby increases soil acidity. The depletion of basic cations in crop harvest, as indicated in their significant reduction is the other cause for the fall in soil pH. The second reason is continuous use of ammonium based fertilizers such as diammonium phosphate. (NH4)₂HPO₄, in such cereal based cultivated fields, which upon its oxidation by soil microbes produces strong inorganic acids. These strong acids in turn

provide H⁺ ions to the soil solution that in turn lower soil pH. Acidic nature of Nitisol was also reported by Yihenew (2002). Thus, it is pertinent to raise the soil pH through liming to increase crop productivity of the study areas.

Due to high amount of rainfall (>1000mm) that results leaching of cations, all of the soil samples had also very low electrical conductivity. The values of exchangeable acidity were also rated as low for most of the sampled soils. A significance (P<0.01) variation in exchangeable acidity was also observed between locations. Exchangeable acidity ranged from 0.15meq/100g at Asendabo to 0.80meq/100g at Serbo (Table 2). The observed small values of exchangeable acidity indicate the easiness to manage the acidity problem of the study areas.

Insert table 2 here

Organic carbon, total nitrogen and available phosphorous

The mean values of organic carbon, total nitrogen and available phosphorous across different locations is given in Table 2. Organic carbon and total nitrogen were significantly (P<0.01) varied across locations. Organic carbon was also showed high variability with in the investigated soil samples (CV=26.57%). It ranged from 1.50% at Serbo to 2.71% at Dedo (Table 2). According to Landon (1991) and Sanchez et al, (1982), all of the sampled soils had organic carbon greater than the critical level (0.5-1%). High organic carbon in Nitisol was also reported by Shimeles et al (2006); Wakene and Heluf (2001); Mesfin, (1998) and Eylachew (1999). The surface soils of Nitisol may contain several percent of organic carbon (WRB, 2006). The highest values of organic carbon could be because of poor drained condition of the soil and high amount of rainfall that reduces the rate of organic matter decomposition in the study sites.

Total nitrogen also showed high variability (CV = 22.87%) among the soil samples. Total nitrogen ranged from 0.16% at Serbo to 0.25% at Dedo. Based on the classification of Landon (1991) and Sanchez et al, (1982), total nitrogen was found as one of the limited plant nutrient in the study sites. The values of total nitrogen in all soil samples were below the critical level (<1%) (Table 2). The observed nitrogen deficiency in all soil samples could be because of low input of plant residues, nitrogen rich organic materials like manure and compost in cereal based farming systems. As the area receives high rainfall, the nitrogen leaching problem can be another reason for the decline of total nitrogen in cropped fields. Moreover, farmers of the study area do not integrate

leguminous plants on their farmlands. Shimeles *et al* (2006) also reported nitrogen deficiency in the cultivated Nitisol.

A significance (P<0.05) difference in available phosphorous was also observed between locations. High variability (CV = 75.31%) in available phosphorus was also observed among the sampled soils. Available phosphorous ranged from 7.19ppm at Agaro to 12.16ppm at Asendabo (Table 2). From the soil samples, 88% had available phosphorous below the critical level (10-15 ppm) given by Landon (1991) and Sanchez et al., (1982). The low level of available phosphorous in the study area might be due to its fixation by Al and Fe, as their presence is expected at the pH values of the soils of the study areas (Tisdale et al., 1993). High phosphorous sorption capacity of Nitisol was also reported by WRB (2006). The correlation matrix (Table 5) also showed a positive and significance (r = 0.37*) relationship between soil pH and available phosphorous. Unlike total nitrogen and available phosphorus, the carbon to nitrogen ratio; however, was with in the critical level except at Serbo which was below the critical level set by Sanchez et al. (1982). About 18% of the sampled soils had C/N ratio less than 10 whereas the rest have C/N ratio with in the critical level.

Cation Exchange Capacity CEC) and exchangeable bases

exchange capacity (CEC) Cation showed significance (P<0.05) difference between locations (Table 3). High variability (CV% = 16.81%) in CEC was also observed among the sampled soils. The value of CEC ranged from 22.97 at Serbo to 28.36 Cmolec/kg at Dedo. The mean value of CEC rated as medium at all study sites except for Dedo which is rated as high (Landon, 1991; Sanchez et al, 1982). From the sampled soils, 32% had the CEC between 25 to 30 Cmolec/Kg while the rest have CEC less than 25 Cmolec/Kg. The relative small amount of CEC in the sampled soil could be because of the amount and nature of the clay. Fasil and Charles (2009), reported that the amount of clay and mainly the type of clay mineral are responsible factors for CEC. The clay assemblage of Nitisol is dominated by kaolinite/(meta) halloysite which has the CEC value of 3-15Cmol(+)/kg (WRB, 2006). The presence of high organic matter in the soil samples; however, contributes to the slight increase in CEC. The correlation matrix (Table 5) also showed a positive and significant (r = 0.59**)relationship between organic carbon and CEC. Moreover, charges from broken edges and exposed OH planes dominate the CEC of kaolinite and charges on kaolinite are pH dependant (Chi and Richard,

Table 3: Mean values of CEC and exchangeable base across different locations in Jimma Zone

Location	Dimtu	Dedo	Asendabo	Agaro	Serbo	Critical level	P-value
CEC (Cmol _c /Kg)	b 24.02	28.36 ^a	b 24.33	b 23.44	22.97 ⁰	_	*
K (Cmol _c /Kg)	1.34 ^b	1.24 ⁰	2.80 ^{ab}	3.54 ^a	1.32 ⁰	<0.2	**
Na (Cmol _c /Kg)	0.22 ^b	0.11 ^b	0.47 ^a	0.26 ^b	0.12 ^b	-	**
Ca (Cmol _c /Kg)	8.08 ^{au}	9.80 ^{ab}	18.58 ^a	6.98 ^b	7.45 ^{ab}	<5.0	NS
Mg (Cmol _c /Kg)	2.86 ^{abc}	3.09 ^{ab}	3.63 ^a	2.62 ^{abc}	1.97 [°]	<1.5	*
PBS (%)	51.41 ^b	49.99 ⁰	70.50 ^a	58.02 ^{ab}	47.97 ⁰	-	*
K (%)	5.57 ^b	4.45 ^b	11.43 ^{ab}	16.21 ^a	5.87 ^b	-	**
Na (%)	0.88 ^{DC}	0.39 ^c	1.73 ^a	1.19 ^{ab}	0.54 ^{DC}	-	**
Ca (%)	33.10	34.26	42.92	29.53	32.95	-	NS
Mg (%)	11.86 ^{ab}	10.90 ^{DC}	14.40 ^a	11.09 ^{DC}	8.61 ^c	-	**

Means in the same row followed by the same letters are not significantly different at 5% level of significance; *significant at P=0.05; **significant at P=0.01; NS = non significance; CEC = cation exchange capacity; PBS = percentage of base saturation

1999). In harmony with this, the correlation matrix (Table 5) of the study showed a positive and significant ($r = 0.58^{**}$) relationship between pH-H₂Oorganic carbon and CEC.

The exchange sites of Nitisol at Jimma zone are mainly occupied by Ca and Mg and to a lesser extent by K and Na. Except exchangeable Ca, all exchangeable bases showed significance difference between locations. High variability in exchangeable bases was also observed among the soil samples. The mean values of exchangeable K ranged from 1.24 at Dedo to 3.54 Cmol_c/Kg at Agaro; exchangeable Ca reanged from 6.98 at Agaro Cmol_c/Kg to 18.58 Cmol_c/Kg at Asendabo; exchangeable Mg ranged from 1.97 Cmol_c/Kg at Serbo to 3.63 Cmol_c/Kg at Asendabo (Table 3). The values of exchangeable K were above the critical level in all sampled soils. However, 4% and 14% of the sampled soils had exchangeable Mg and Ca, respectively below the critical level given by Landon (1991) and Sanchez et al., (1982).

A significant (P<0.05) difference in percentage of base saturation was also observed between locations. High variation (CV = 30.23%) in percentage of bases saturation was also observed among the soil samples. According to Landon (1991) and Sanchez et al, (1982), the percentages of base saturation for all soil samples are rated as medium to high.

Insert table 3 here

Micronutrient Status

Micronutrient status of Nitisol in Jimma Zone is presented in Table 4. Micronutrient status of soils of the study areas showed a significant (P<0.01) different between locations. High variability in micronutrient

status was also observed among the soil samples. The availability of trace elements in the soil is influenced by many soil and environmental factors (Jones, 1972). According to most literatures, the availability of micronutrients increases in acidic soil (Vijay et al., 1997); however, according to the critical levels provided by Jones and Eck (1973), only available Mn and Fe were found above the critical level for all sampled soils. However, all soil samples had the concentration of available Cu and Zn below the critical value provided by Jones and Eck (1973). The correlation matrix (Table 5) also showed a non significance relationship between micronutrients and soil pH. Higher crop yields which increase plant nutrient demands, use of fertilizers containing lower auantities of micronutrient contaminants. decreased use of farmyard manure results low Cu and Zn concentration in the sampled soils. Deficiency of micronutrients in Ethiopian soils was reported by (Louis, 2010; Teklu et al., 2007 and Wakene and Heluf, 2001).

Insert table 4 and 5 here

Nitrogen and Phosphorous Fertilizer Recommendation using NuMaSS

The recommended amounts of nitrogen and phosphorous for maize across different locations using NuMaSS tool are presented in Table 5. According to the tool, high probabilities of phosphorous and nitrogen deficiencies were observed in all study sites. The recommended amount of phosphorous for maize is ranged from 4.97PKg/ha at Asendabo to 27Kg P/ha at Dimtu while the recommended amount of N is ranged from 73 Kg/ha at Dedo to 100Kg/ha at Serbo. The

Table 4: Mean values of micronutrients across different locations in Jimma zone

Location	Mn	Fe	Cu	Zn	
(ppm)					
Dimtu	b 102.17	b 45.80	0.40 b	C 1.98	
Dedo	101.88 ⁰	54.51 ⁰	0.53 ^b	4.73 ^{ab}	
Asendabo	a 114.06	a 71.95	a 0.84	ab 4.58	
Agaro Serbo	121.41 ^a	52.55 ⁰	0.94 ^a	5.24 ^a	
Serbo	97.07	53.50 b	0.51	b 3.54	
Critical level	50.00	20.00	4.00	20.00	
P-Value	**	**	**	**	

Means in the same column followed by the same letters are not significantly different at 5% level of significance; *significant at P<0.05; **significant at P<0.01

Table 5: Pearson's correlation matrix for the selected soil fertility parameters

	Clay	pH-H ₂ O	OC	TN	AVP	CEC	K
Clay	-						
pH-H ₂ O	0.13	-					
OC _	0.09	0.37*	-				
TN	0.36*	0.17	0.77**	-			
AVP	0.51**	0.37*	-0.02	-0.10	-		
CEC	0.33	0.58*	0.59**	0.57**	0.34	-	
K	0.51**	0.13	0.18	0.13	0.25	-0.01	-
Na	0.32	0.53**	0.26	0.19	0.39*	0.41*	0.63
Mg	0.18	0.58**	0.39*	0.39*	0.33	0.72**	0.19
Ca	0.23	0.48**	0.19	0.18	0.45*	0.59**	0.16
PBS		0.23	0.02	-0.01	0.37*	0.08	0.69**
Mn	0.21	-0.22	0.12	0.18	0.10	-0.15	0.52**
Fe	0.12	0.12	-0.13	-0.18	0.16	-0.23	0.20
Zn	0.17	-0.23	0.12	0.10	-0.04	-0.16	0.48**
Cu	0.08	-0.17	0.08	-0.10	0.01	-0.19	0.40*

^{*} Significant at P<0.05; ** significant at P<0.01; OC = organic carbon; TN = total nitrogen; AVP = available phosphorus; CEC = cation exchange capacity

Table 6: Nitrogen and phosphorous fertilizers recommendation for maize across different locations using NuMaSS

	Maize			
	Prob.	Recommended P (Kg/ha)	Prob.	Recommended N (Kg/ha)
	Level (%)		Level (%)	
Dimtu	92	27	87	94
Dedo	91	24	87	73
Asendabo	85	4.9	87	97
Agaro	91	24	87	82
Serbo	90	20	87	100

smallest phosphorous recommendation at Asendabo could be because of the residual effect of fertilization. Due to its proximity to the city, the intensive agricultural practices demand excess application of DAP at Asendabo. In line with NuMaSS fertilizer recommendation, the blanket fertilizer recommendation in the study areas for maize is 100kg DAP and 100Kg urea which is equivalent to 20.08 Kg/ha of phosphorous and 64 Kg/ha of nitrogen. Moreover, 110

Kg N and 20 Kg P was recommended for maize in Nitisol (Wakene et al., 2007). Therefore, NuMaSS could be used as a tool to diagnose and recommend the amount of fertilizer required for maize in different agroecology of the study area.

Insert table 6 here

CONCLUSION AND RECOMMENDATION

The study revealed that the fertility status of Nitisol is highly varied across location and it has serious nutrient limitations. All of the sampled soils have pH-H₂O below the critical limit for plant growth. Nitrogen, Cu and Zn deficiency was also observed in all soil samples. Similarly, 88% of the sampled soils have available phosphorous below the critical level. Therefore, management practices such as increase input of plant residues, nitrogen rich organic materials like manure and compost are required. Moreover, application of lime, micronutrients based fertilizers should be introduced in order to boost the crop productivity of the study area. Since the fertilizer recommendation from field experiment is in harmony with NuMaSS fertilizer recommendation, NuMaSS can be used as a tool in diagnose nutrient deficiency order to recommended fertilizers for maize in different agroecology. Finally, for validation, it is recommended that the study should be repeated in different soils and agroecological zones in Ethiopia and elsewhere.

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REFERENCES

- Bojo J, Cassels D (1995). Land Degradation and Rehabilitation in Ethiopia: A Reassessment. World Bank Africa Technical Department Working Paper 17. Washington, D.C.
- BPEDORS (2000). Physical and Socio-Economical Profile of 180 District of Oromia Region. Bureau of Planning and Economic Development, Council of Regional State of Oromiya. Ethiopia
- Campbell CA, McConkey BG, Zentner RP, Sells F, Curtin D (1996). Long-term effects of tillage and crop rotations on soil organic carbon and total nitrogen in a clay soil in south western Saskatchewan. Canadian Journal of Soil Sciences. 76: 395-340.
- Chapman HD (1965). Cation Exchange Capacity. In: C.A. Black, *et al.* (Eds.), Methods of Soil Analysis. Agronomy 9, American Society of Agronomy. Inc., Madison, Wisconsin, pp. 891–901.
- Chi M, Richard AE (1999). Cation Exchange Capacity of Kaolinite. Clays and Clay Minerals' 47 (2): 174-180.
- Elias E (2002). Farmers Perception of Soil Fertility Change and Management. SOS-Sahel and Institute for sustainable Development, Addis Ababa, Ethiopia, pp: 252
- Eylachew Z (1999). Selected physical, chemical and mineralogical characteristics of major soils occurring in Chercher highlands, eastern Ethiopia. Ethiopian Journal of Natural Resource,1(2): 173-185.
- Fassil K and Charles Y (2009). Soil Fertility Status and Numass Fertilizer Recommendation of Typic Hapluusterts in the Northern

- Highlands of Ethiopia. World Applied Sciences Journal 6 (11): 1473-1480.
- Hurni H (1988). Degradation and conservation of the resources in the Ethiopian highlands. Mountain Research and Development, 8 (2/3): 123-130.
- Jones J, Eck HV (1973). Plant analysis as an aid in fertilizing corn and grain sorghum. In Soil Testing and Plant Analysis and Revised L.M. Walsh and J.D. Beaton (eds.), Soil Science Society of American, pp. 349-364.
- Jones JB (1972). Plants tissue analysis for micronutrients, In: "Micronutrients in Agriculture (Ed. Mortvedt J.J., Giodano P.M., Lindsay W.L.), Soil Science Society of America, Madison, Wisconsin. pp. 319.
- Landon JR (1991). Booker Tropical Soil Manual, A handbook of soil Survey and Agricultural Land evaluation in the tropics and subtropics. Longman publication. London. pp 500.
- Lindsay WL, Norvell WA (1978). Development of a DTPA soil test for zinc, iron, manganese and copper. Soil Science Society of American Journal. 42:421-428.
- Louis EA (2010). Physicochemical characteristics of the rhizosphere soils of some cereal crops in Ambo Woreda, West Shoa, Ethiopia. Maejo International Journal of Science and Technology. 4(1), 93-100.
- Mesfin A (1998). Nature and management of Ethiopian soils. Alemaya University of Agriculture, Ethiopia. pp 272.
- Quinones, MA, Borlaug NE, Dowswell CR (1998). A fertilizer-based green revolution in Africa. In: RJ Buresh, PA Sanchez & F Calhoun (eds), Replenishing soil fertility in Africa. SSSA, Special Publication No.51, Madison, WI, USA. pp 81-95.
- Sanchez PA, Couto W, Boul SW (1982). The fertility capability soil classification system: interpretation, applicability and modification. Geoderma, 27: 83-309.
- Sánchez PA, Valencia I, Izac AM, Pieri C (1995). Soil fertility replenishment in Africa. Nairobi, Kenya: ICRAF.
- Shapiro BI, Sanders JH (1998). Fertilizer use in semiarid West-Africa. Profitability and supporting policy. Agricultural Systems, 56 (5): 467–482.
- Shepherd KD., Soule MJ (1998). Soil fertility management in West Kenya: Dynamic simulation of productivity, profitability and sustainability at different resources endowment levels. Agric. Ecosystystem, Environment, 71: 133-147.
- Shimeles D, Mohammed A, Abayneh E (2006). Characterization and Classification of the Soils of Tenocha-Wenchancher Micro Catchment, Southwest Shewa. M.Sc thesis, Alemaya University, Ethiopia. pp 111.
- Stoorvogel JJ, Smaling EMA (1990). Assessment of soil nutrient depletion in Sub-Saharan Africa: 1983-2000. Report 28, DLO Winand Staring Center for Integrated Land, Soil and Water Research (SC-DLO), Wageningen, Netherlands.
- Taye B, Yifru A (2010). Assessment of Soil Fertility Status with Depth in Wheat Growing Highlands of Southeast Ethiopia. World Journal of Agricultural Sciences 6 (5): 525-531.
- Teklu B, Amnat S, Yongyuth O, Ed S (2007). Status of Mn, Fe, Cu, Zn, B and Mo in Rift Valley Soils of Ethiopia: Laboratory Assessment. Kasetsart Journal (Nat. Sci.) 41: 84 95.
- Tisdale SL, Nelson WL, Beaton JD, Havlin JL (1993). Soil fertility and fertilizers. 5th edition. Macmillan Publishing Co., NY, pp 634.
- Van Reewijk LP (1992). Procedures for Soil Analysis (3rd Ed.). International Soil Reference Center (ISRIC), Wageningen, The Netherlands, pp 371.
- Vijay MV, Bruce RJ, Alba T (1997). Copper solubility in Myersville B horizon soil in the presence of DTPA. Soil Science Society of American Journal. 61: 44-52.
- Wakene N, Heluf G (2001). Assessment of Important Physicochemical Properties of Dystric Udalf (Dystric Nitosols) under different Management Systems in Boko Area, Western Ethiopia. MSc Thesis, Alemaya University, Ethiopa. pp 93
- Wakene N, Tolera A, Friesen DK, Abdenna D, Berhanu D (2001). Evaluation of compost for maize production under farmers'

- condition. Seventh Eastern and Southern Africa Regional Maize Conference. 11th-15th February, pp 382-386.

 Weigel G (1986). The soils of the Gununo area, Sidamo Research Unit, Ethiopia. Research report 8, Soil Conservation Research Project, Addis Ababa. pp 94.
- World Reference Base (WRB) (2006). World Reference Base for Soil Resources: A framework for International Classification,
- Correlation and Communication. Food and Agriculture Organization of the United Nations. Rome, Italy. pp 145.
- Yihenew G (2002). Selected physical and chemical characteristics of soils of Adet Research Center and its testing sites in northwestern Ethiopia. Ethiopian Journal of Natural Resources 4(2): 199-215